PREPARATION OF ANDROSTANES RELATED TO APHIDICOLIN

Cavit UYANIK*a,**, Kudret YILDIRIM*^b*, Aslihan MALAY*^c* , James R. HANSON*d1* and Peter B. HITCHCOCK*d2*

- *^a Department of Chemistry, University of Kocaeli, Izmit, 41380, Kocaeli, Turkey; e-mail: cuyanik@kou.edu.tr*
- *^b Department of Chemistry, University of Sakarya, Adapazari, 54187, Sakarya, Turkey; e-mail: kudrety@sakarya.edu.tr*
- *^c Department of Chemistry, Gebze Institute of Technology, Gebze, 41400, Kocaeli, Turkey; e-mail: amalay@gyte.edu.tr*
- *^d Department of Chemistry, University of Sussex, Brighton, Sussex, BN1 9QJ, U.K.; e-mail: ¹ j.r.hanson@sussex.ac.uk, ² p.b.hitcock@sussex.ac.uk*

Received July 25, 2007 Accepted November 9, 2007

The preparation of 3α,4α,17β-trihydroxy-, 3α,4α,16α,17α- and 2α,3α,16α,17α-tetrahydroxy-5α-androstane derivatives (**5**, **11**, **18**) from testosterone as steroidal analogues of diterpenoid inhibitor of DNA polymerase α , aphidicolin, is described. The crystal structure of 5α-androstane-3α,4α,17β-triyl triacetate (**6**) is reported.

Keywords: Aphidicolin; Steroids; Androstanes; Diterpenoids; Catalytic hydroxylation; X-ray crystallography; DNA polymerase inhibitors.

Aphidicolin (1) is an inhibitor of DNA polymerase α (refs^{1,2}). Its glycinate ester reached clinical trial as a tumour inhibitory substance. The structureactivity studies which have been reported 3 on this relatively expensive diterpenoid suggest that the activity may be related to the distance separating the hydroxyl groups on rings A and D of aphidicolin. There is a formal similarity between the structure of aphidicolin and the steroids and, unlike the majority of the diterpenoids, both belong to the same enantiomeric series. The binding of the steroid hormones to the steroid nuclear receptors results in the initiation of nucleic acid biosynthesis. This involves DNA polymerase activity. Consequently, we have prepared $4-6$ a number of steroids with structures that have a similarity to aphidicolin in the relative disposition of their hydroxyl groups. The C-3:C-17 and C-4α (hydroxymethyl): C-17 distances in aphidicolin are 8.46 and 8.84 Å whilst in a typical 5α steroid C-3:C-17 is 8.70 and C-4:C-17 8.09 Å. Preliminary screening results suggest that some of these compounds possess tumour inhibitory activity.

1546 Uyanik, Yildirim, Malay, Hanson, Hitchcock:

In this paper we report the preparation of 5α-androstane-3α,4α,17βtriol (**5**), 5α-androstane-3α,4α,16α,17α-tetrol (**11**) and 5α-androstane-2α,3α,16α,17α-tetrol (**18**) from the readily available testosterone (**2**). These compounds have a 3α-hydroxyl group typical of aphidicolin.

5α-Androst-3-en-17β-yl acetate⁷ (**3**) was obtained from testosterone (**2**) by treatment with boron trifluoride etherate-sodium borohydride and acetic anhydride. Hydrolysis of the 17β-acetate gave the 17β-alcohol⁸ **4**. Catalytic osmylation of 5α-androst-3-en-17β-ol (**4**) using potassium hexacyanoferrate(III) as the co-oxidant⁹, gave 5α-androstane-3α, 4α, 17βtriol (**5**) (Scheme 1). The 1H NMR spectrum of the product, determined in pyridine- d_5 at 500 MHz, contained signals at δ 3.69 (1 H, dd, $J = 3$ and

SCHEME 1

11 Hz, 4-H) and δ 4.28 (1 H, q, $J = 3$ Hz, 3-H). The magnitude of the coupling constants was consistent with the presence of the 4α - and 3α -hydroxyl groups, respectively. Thus the 4-H resonance revealed a diaxial and an axial-equatorial coupling whilst the 3-H resonance contained equatorialaxial and diequatorial couplings. The stereochemistry was confirmed by an X-ray crystal structure of the triacetate **6** as shown in Fig. 1.

Oxidation of 5α-androst-3-en-17β-ol (**4**) with chromium trioxide gave 5α-androst-3-en-17-one⁸ (**7**) which was converted to its hydrazone **8** with hydrazine hydrate and triethylamine. Decomposition of the hydrazone with iodine and triethylamine in tetrahydrofuran¹⁰ gave 17-iodo-5α-androsta-3,16-diene (**9**). Apart from the signals assigned to the ring A double bond, the ¹H NMR spectrum of the product contained a new alkene signal at δ 6.12. Hydrogenolysis of the iodine with sodium in ethanol afford 5α-androsta-3,16-diene (**10**) which possessed signals of olefinicprotons at δ 5.69 and 5.84 (H-17 and H-16, respectively) and at δ 5.26 and 5.52 (H-4 and H-3, respectively). This method of preparing the 16-enes is preferable over that of dehydration of the 17-alcohols which can bring about rearrangement of C-18.

Catalytic osmylation of diene **10** gave 5α-androstane-3α,4α,16α,17αtetrol (11). The ¹H NMR spectrum determined in pyridine- d_5 at 500 MHz, contained signals at δ 3.67 (doublet of doublet, $J = 3$ and 11 Hz), δ 3.82 (doublet, $J = 5.3$ Hz), δ 4.27 (quartet, $J = 3$ Hz) and δ 4.67 (quartet, $J =$

FIG. 1 X-ray structure of **6**

5.3 Hz) which were assigned to H-4, H-17, H-3 and H-16, respectively. Irradiation of the H-18 signal at δ 0.70 gave nuclear Overhauser effect enhancements of 6.4% to the H-17 signal and 4.4% to the H-16 signal, showing that both the 16- and 17-hydroxyl groups were α -oriented. Irradiation of the H-19 signal at δ 0.85 gave an NOE enhancement of 4.4% to the H-4 signal at δ 3.67. These enhancements, together with the magnitude of the coupling constants, were consistent with the 5α-androstane-3α,4α,16α,17α-tetrol stereochemistry of the compound **11**.

5α-Androstane-2α,3α,16α,17α-tetrol (**18**) was prepared by a similar route (Scheme 2) from 3β-hydroxyandrost-5-en-17-one (**12**). 5α-Androst-2-en-17-one¹¹ (**14**) was obtained by elimination of the 3β-tosylate **13** with collidine and converted into 5α-androsta-2,16-diene (**17**) via the 17 hydrazone **15** and the 17-iodo-2,16-diene (**16**) as above. Catalytic osmylation of **17** gave the tetrol **18**. The 1H NMR spectrum contained signals at δ 3.80 (doublet, *J* = 5 Hz), δ 4.01 (multiplet), δ 4.30 (quartet *J* = 3 Hz) and δ 4.66 (multiplet) which were assigned to H-17, H-2, H-3 and H-16, respectively. Irradiation of the H-18 signal at δ 0.67 gave an NOE enhancements of 4.7% to the signal at δ 3.80 and 3.4% to the signal at δ 4.66, whilst irradiation of the H-19 signal at δ 0.82 gave an NOE enhancement of 2.6% to

SCHEME 2

the signal at δ 4.01. This, together with multiplicity of the H-3 signal, showed that the tetrol was the 5α-androstane-2α,3α,16α,17α-tetrol (**18**).

In conclusion the syntheses of 5α-androstane-3α,4α,17β-triol, 5α-androstane-3α,4α,16α,17α-tetrol, 5α-androstane-2α,3α,16α,17α-tetrol as aphidicolin models were described. The stereochemistry of osmylation followed by acetylation of 5α-androst-3-en-17β-ol was confirmed by an X-ray crystallography.

EXPERIMENTAL

Melting points were determined using an Electrothermal IA 9200 apparatus and are uncorrected. ¹H NMR spectra were recorded in deuteriochloroform and pyridine- d_5 with tetramethylsilane as an internal standard reference at 300 and 500 MHz on a Bruker DPX 300 and Bruker AMX 500 spectrometers. 13 C NMR spectra were recorded in deuteriochloroform at 75 and 100 MHz on a Bruker DPX 300 and Bruker AMX 500 spectrometers. Chemical shifts are given in ppm (δ-scale), coupling constants (*J*) are given in Hz. IR spectra (wavenumbers in cm^{-1}) were recorded using Nujol mulls on a Perkin–Elmer 1710 Fourier transform spectrometer. High-resolution mass spectra (HRMS) were determined on a Bruker Daltonics Apex III mass spectrometer operating in the electrospray mode. The X-ray analysis data collection was performed using KappaCCD. The refinements were performed using SHELXL-97 and the figures were drawn using ORTEP-3 for Windows. A stock solution of osmium tetraoxide (1 g, 4 mmol) in redistilled *tert*-butanol (80 ml) containing *tert*-butyl hydroperoxide (2 ml) was prepared and stored at 0 °C. Extracts were dried over anhydrous sodium sulfate. Light petroleum refers to the fraction of b.p. 60–80 °C. Silica gel for chromatography was Merck 9385.

5α-Androst-3-ene-17-one (**7**)

This compound was prepared according to the literature procedure⁸. Yield 82%, m.p. 120–122 °C (lit.⁸ gives m.p. 121–123 °C). IR: 1725 (C=O). ¹H NMR (300 MHz, CDCl₃): 0.80 s, 3 H (H-18); 0.88 s, 3 H (H-19); 5.26 dd, 1 H, *J*(4,5) = 2.0, *J*(4,3) = 9.7 (H-4); 5.52 dq, 1 H, $J(3,2) = 3.0, J(3,4) = 9.7$ (H-3).

5α-Androst-3-en-17-one Hydrazone (**8**)

Ketone⁸ **7** (300 mg, 1.1 mmol) in dry tetrahydrofuran (15 ml) was heated under reflux with triethylamine (0.58 ml, 4.2 mmol) and hydrazine hydrate (2 ml, 99%) for 3 h. The solution was poured into cold water (20 ml) and the mixture cooled in ice. The precipitate was collected and washed with water to give the hydrazone **8** (290 mg, 92%) which was crystallized from ethyl acetate as plates, m.p. 160-163 °C. For C₁₉H₃₀N₂ (286.4) calculated: 79.67% C, 10.56% H, 9.78% N; found: 79.59% C, 10.48% H, 9.67% N. IR: 3172 (N-H). ¹H NMR (300 MHz, CDCl3): 0.70 s, 3 H (H-18); 0.88 s, 3 H (H-19); 4.74 bs, 2 H (NH2); 5.26 dd, 1 H, *J*(4,5) = 2.0, $J(4,3) = 9.7$ (H-4); 5.52 dq, 1 H, $J(3,2) = 3.0$, $J(3,4) = 9.7$ (H-3). ¹³C NMR (75 MHz, CDCl₃): 166.4, 131.2, 125.3, 53.7, 53.6, 45.9, 44.2, 35.0, 34.9, 34.2, 34.0, 31.4, 27.2, 24.3, 23.4, 23,3, 20.7, 17.1, 11.8.

5α-Androst-2-en-17-one Hydrazone (**15**)

Under similar conditions 5α-androst-2-en-17-one¹¹ (**14**; 300 mg, 1.1 mmol) gave the hydrazone **15** (300 mg, 95%) as a white solid, m.p. 158-160 °C. For $C_{19}H_{30}N_2$ (286.4) calculated: 79.72% C, 10.69% H, 9.80% N; found: 79.70% C, 10.63% H, 9.28% N. IR: 3170 (N-H), 1630 (C=C). ¹H NMR (300 MHz, CDCl₃): 0.78 s, 3 H (H-18); 0.86 s, 3 H (H-19); 4.75 bs, 2 H (NH₂); 5.55 m, 2 H (H-2 and H-3). ¹³C NMR (75 MHz, CDCl₃): 166.4, 125.8 (2 × C); 54.3, 53.6, 43.9, 41.4, 39.67, 34.9, 34.7, 34.2, 31.2, 30.2, 28.5, 24.3, 23.3, 20.5, 16.9, 11.6.

17-Iodo-5α-androsta-3,16-diene (**9**)

The hydrazone **8** (300 mg,1.05 mmol) and triethylamine (1.4 ml, 10 mmol) in dry tetrahydrofuran (15 ml) was treated with a solution of iodine (400 mg, 1.6 mmol) in tetrahydrofuran (5 ml) at room temperature under nitrogen. After the evolution of nitrogen had ceased, the solvent was evaporated and the residue was taken up in ethyl acetate. The extract was washed with dilute hydrochloric acid, water, aqueous sodium sulfite, aqueous sodium hydrogen carbonate and brine, and dried. The solvent was evaporated and the residue chromatographed on silica gel. Elution with light petroleum gave the diene **9** (320 mg, 80.5%) as a gum. IR: 1659 (C=C). ¹H NMR (300 MHz, CDCl₃): 0.74 s, 3 H (H-18); 0.80 s, 3 H (H-19); 5.26 dd, 1 H, *J*(4,5) = 2.0, *J*(4,3) = 9.7 (H-4); 5.52 dq, 1 H, *J*(3,2) = 3.0, *J*(3,4) = 9.7 (H-3); 6.12 d, 1 H, *J* = 1.2 (H-16). ¹³C NMR (75 MHz, CDCl₃): 137.4, 131.0, 125.5, 113.0, 54.5, 53.6, 50.1, 45.9, 36.2, 35.0, 34.6, 33.8, 33.7, 31.5, 27.2, 23.4, 20.8, 15.3, 11.8. HRMS: for $C_{10}H_{27}$ I calculated: 382.1150; found: 382.1148.

17-Iodo-5α-androsta-2,16-diene (**16**)

Under similar conditions the hydrazone **15** (280 mg, 0.97 mmol) gave the diene **16** (300 mg, 80%) as a gum. IR: 1659 (C=C). ¹H NMR (300 MHz, CDCl₃): 0.74 s, 3 H (H-18); 0.78 s, 3 H $(H-19)$; 5.56 m, 2 H (H-2- and H-3); 6.12 d, 1 H, $J = 1.2$ (H-16). ¹³C NMR (75 MHz, CDCl₃): 137.4, 125.8 $(2 \times C)$; 112.9, 54.7, 54.3, 49.9, 41.5, 39.5, 36.2, 34.8, 34.6, 33.6, 31.3, 30.2, 28.5, 21.6, 15.2, 11.6. HRMS: for $C_{19}H_{27}I$ calculated: 382.1162; found: 382.1180.

5α-Androsta-3,16-diene (**10**)

The above iodoalkene **9** (250 mg, 0.6 mmol) in ethanol (40 ml) was treated with sodium (2 g, 87 mmol) under reflux for 20 min. The solution was cooled and treated with 50% aqueous ethanol (40 ml). The ethanol was evaporated in vacuo and the residue extracted with ethyl acetate. The extract was washed with dilute hydrochloric acid, aqueous sodium sulfite, water and brine, and dried. The solvent was evaporated and the residue was chromatographed on silica gel. Elution with light petroleum gave the diene **10** (140 mg, 83.5%) as an oil. IR: 1650 (C=C). ¹H NMR (300 MHz, CDCl₃): 0.76 s, 3 H (H-18); 0.78 s, 3 H (H-19); 5.26 dd, 1 H, *J*(4,5) = 2.0, *J*(4,3) = 9.7 (H-4); 5.52 dq, 1 H, *J*(3,2) = 3.0, *J*(3,4) = 9.7 (H-3); 5.69 bs, 1 H (H-17); 5.84 dd, 1 H, $J(16,15) = 1.8$, $J(16,17) = 4.7$ (H-16). ¹³C NMR (75) MHz, CDCl₃): 143.9, 131.4, 129.5, 125.4, 56.2, 54.2, 46.1, 45.7, 35.9, 35.2, 34.2, 33.9, 32.1, 29.7, 27.4, 23.5, 21.0, 17.1, 11.9. HRMS: for C₁₉H₂₈ calculated: 256.2191; found: 256.2200.

5α-Androsta-2,16-diene (**17**)

Under similar conditions the iodoalkene **16** (250 mg, 0.6 mmol) gave the diene **17** (135 mg, 80.5%) as an oil. IR: 1640 (C=C). ¹H NMR (300 MHz, CDCl₃): 0.78 s, 3 H (H-18); 0.80 s, 3 H (H-19); 5.56 m, 2 H (H-2 and H-3); 5.69 d, 1 H, $J(17,16) = 4.7$ (H-17); 5.84 dd, 1 H, $J(16,15) =$ 1.8, $J(16,17) = 4.7$ (H-16). ¹³C NMR (75 MHz, CDCl₃): 143.9, 129.2, 125.9 (2 × C); 56.1, 54.8, 45.5, 41.6, 39.6, 35.9, 34.9, 34.2, 31.9, 31.8, 30.3, 28.7, 20.8, 17.0, 11.6. HRMS: for $C_{19}H_{28}$ calculated: 256.2195; found: 256.2200.

5α-Androstane-3α,4α,17β-triol (**5**)

5α-Androst-3-en-17β-ol⁸ (**4**; 140 mg, 0.5 mmol) in *tert*-butanol (15 ml) and water (15 ml) was treated with potassium hexacyanoferrate(III) (1.65 g, 5 mmol), potassium carbonate (690 mg, 5 mmol), 1,4-diazobicyclo[2,2,2]octane (56 mg, 0.5 mmol) and the stock solution of osmium tetraoxide (0.4 ml) at 40 °C for 24 h. Sodium sulfite $(140 \text{ mg}, 1.1 \text{ mmol})$ was added and the solution was then left to stir overnight. The mixture was filtered and extracted with ethyl acetate. The extract was dried and the solvent was evaporated to give the triol **5** (104 mg, 71%) which was crystallized from ethyl acetate as needles, m.p. 194–196 °C. For $C_{19}H_{32}O_3$ (308.5) calculated: 74.00% C, 10.52% H; found: 73.65% C, 10.37% H. IR: 3490 (O-H). ¹H NMR (500 MHz, pyridine-*d*₅): 0. 87 s, 3 H (H-18); 0.96 s, 3 H (H-19); 3.69 dd, 1 H, *J* = 3.0, 11.0 (H-4); 3.83 t, 1 H, *J* = 8.5 (H-17); 4.28 q, 1 H, *J* = 3.0 (H-3). 13C NMR (100 MHz, pyridine-*d*5): 80.6, 71.1, 69.4, 55.2, 50.7, 45.4, 42.7, 36.9, 36.8, 34.8, 31.6, 31.1, 29.9, 27.3, 23.0, 22.5, 20.2, 12.3, 11.3. The triacetate **6**, prepared with acetic anhydride in pyridine, had m.p. 184–186 °C. IR: 1745 (C=O, acetate), 1716 (C=O, acetate). ¹H NMR (300 MHz, CDCl₃): 0.75 s, 3 H (H-18); 0.85 s, 3 H (H-19); 1.96 s, 3 H (17β-OAc); 2.0 s, 3 H (4α-OAc); 2.06 s, 3 H (3α-OAc); 4.5 t, 1 H, *J* = 8.5 (H-17); 4.7 dd, 1 H, *J* = 3.0, 11.0 (H-4); 5.22 q, 1 H, *J* = 3.0 (H-3). 13 C NMR (75 MHz, CDCl₃): 171.65, 171.17, 170.89, 83.15, 72.71, 69.67, 54.49, 51.11, 44.22, 42.89, 37.89, 37.21, 35.17, 32.34, 31.13, 27.89, 25.61, 23.81, 22.65, 21.65, 21.57, 21.34, 20.71, 13.09, 12.50. HRMS: for $C_{25}H_{38}O_6$ + Na calculated: 457.2560; found: 457.2562.

5α-Androstane-3α,4α,16α,17α-tetrol (**11**)

Under similar conditions the diene **10** (130 mg, 0.5 mmol) gave the tetrol **11** (100 mg, 61%) as a white powder, m.p. 234-236 °C. For $C_{19}H_{32}O_4 \cdot 0.5H_2O$ (333.5) calculated 68.41% C, 10.00% H; found: 68.37% C, 9.95% H. IR: 3470 broad (O-H). 1H NMR (500 MHz, pyridine- d_5): 0. 70 s, 3 H (H-18); 0.85 s, 3 H (H-19); 3.67 dd, 1 H, $J = 3.0$, 11.0 (H-4); 3.82 d, 1 H, *J* = 5.3 (H-17); 4.28 q, 1 H, *J* = 3.0 (H-3); 4.67 q, 1 H, *J* = 5.3 (H-16). 13C NMR (100 MHz, pyridine- d_5): 78.9, 71.6, 71.3, 69.7, 54.6, 47.6, 46.2, 45.2, 37.5, 36.1, 35.5, 32.6, 32.3, 32.1, 28.1, 23.4, 20.3, 17.5, 12.9.

5α-Androstane-2α,3α,16α,17α-tetrol (**18**)

Under similar conditions the diene **17** (130 mg, 0.5 mmol) gave the tetrol **18** (90 mg, 55%) as a white powder, m.p. 220–223 °C. IR: 3450 broad (O-H). ¹H NMR (500 MHz, pyridine- d_5): 0.67 s, 3 H (H-18); 0.82 s, 3 H (H-19); 3.82 d, 1 H, *J*(16,17) = 5.0 (H-17); 4.01 m, 1 H (H-2); 4.30 q, 1 H, $J = 3.0$ (H-3); 4.67 m, 1 H (H-16). ¹³C NMR (100 MHz, pyridine- d_5): 78.9, 71.6, 71.2, 69.8, 54.7, 47.6, 45.6, 41.0, 39.8, 36.3, 36.1, 35.7, 32.6, 32.3, 30.8, 28.2, 20.3, 18.1, 12.9. HRMS: for $C_{19}H_{32}O_4$ calculated: 324.2301; found: 324.2298.

X-ray Crystallographic Data and Structure Determination of 5α-Androstane-3α,4α,17β-triyl Triacetate (**6**)

 $C_{25}H_{38}O_6$, *M_r* 434.55, orthorhombic, space group $P2_12_12_1$ (No. 19), $a = 7.4107(2)$ Å, $b =$ 10.4524(4) Å, *c* = 30.6454(13) Å, α = β = γ = 90°, *V* = 2373.78(15) Å³, Z = 4, D_{cal} = 1.22 g cm⁻³, $\mu = 0.09$ mm⁻¹, F(000) = 944. Data were collected on a crystal of size $0.10 \times 0.02 \times 0.02$ mm³ on a KappaCCD diffractometer operating for $3.83 \le \theta \le 25.1^\circ$. Reflections of 9275 were collected for –7 ≤ *h* ≤ 8, –12 ≤ *k* ≤ 11, –36 ≤ *l* ≤ 29. There were 4057 independent reflections with 2971 possessing $I > 2\sigma(I)$. The structure was solved by direct methods and refined using SHELXL97. The final *R* indices were $[I > 2\sigma(I)]$ $R_1 = 0.057$, $wR_2 = 0.115$ and (all data) $R_1 =$ 0.093 and $wR_2 = 0.133$. The largest difference peak and hole were 0.18 and -0.19 e \AA^{-3} .

CCDC 249909 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge, CB2 1EZ, UK; fax: +44 1223 336033; or deposit@ccdc.cam.ac.uk).

REFERENCES

- 1. Dalziel W., Hesp B., Stevenson K. M., Jarvis J. A. J.: *J. [Chem.](http://dx.doi.org/10.1039/p19730002841) Soc., Perkin Trans. 1* **1973**, [2841.](http://dx.doi.org/10.1039/p19730002841)
- 2. Ikegami S., Taguchi T., Ohashi M.: *[Nature](http://dx.doi.org/10.1038/275458a0)* **1978**, *275*, 458.
- 3. Hiranuma S., Shimizu T., Yoshioka H., Ona K., Nakane H., Takahashi T.: *Chem. Pharm. Bull. Jpn.* **1987**, *35*, 1641.
- 4. Boynton J. A., Hanson J. R.: *J. [Chem.](http://dx.doi.org/10.1039/p19950002189) Soc., Perkin Trans. 1* **1995**, 2189.
- 5. Hanson J. R., Hitchcock P. B., Uyanik C.: *J. Chem. Res., [Synop.](http://dx.doi.org/10.1039/a804002h)* **1998**, 682.
- 6. Hanson J. R., Yildirim K.: *J. Chem. Res., [Synop.](http://dx.doi.org/10.1039/a905656d)* **1999**, 698.
- 7. Cambie R. C., Rutledge P. S., Scott D. W., Woodgate P. D.: *Aust. J. Chem.* **1979**, 695.
- 8. Ihara M., Sudow J., Fukumoto K., Kametani T.: *J. [Chem.](http://dx.doi.org/10.1039/p19860000117) Soc., Perkin Trans. 1* **1986**, 117.
- 9. Minato M., Yamamoto K., Tsuji J.: *J. Org. [Chem.](http://dx.doi.org/10.1021/jo00289a066)* **1990**, *55*, 766.
- 10. Barton D. H. R., O'Brian R. E., Sternhell S.: *J. [Chem.](http://dx.doi.org/10.1039/jr9620000470) Soc.* **1962**, 470.
- 11. Knox L. H., Velarde E., Berger S., Cuadriello D., Cross A. D.: *J. Org. [Chem.](http://dx.doi.org/10.1021/jo01031a019)* **1964**, *29*, [2187.](http://dx.doi.org/10.1021/jo01031a019)